



***Khaya senegalensis* FLAVONOID MITIGATE ALUMINUM CHLORIDE-INDUCED
ISCHEMIC STROKE IN THE PARIETOTEMPORAL CORTEX IN WISTAR
RATS**

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Abstract

Restorative approaches to treat stroke is still the major challenges confronting stroke researches in attempt to manage and treat stroke. This study investigated ameliorative effects of *Khaya senegalensis* flavonoids on aluminum chloride-induced ischemic stroke in the parieto-temporal cortex. Thirty-five healthy adult male rats weighing 150–220g, were randomly assigned into five groups (n =6): Group 1: serve as control received 1 mg/kg of distilled water, Group 2 (AlCl₃ -2Wks)i, received 100 mg/kg of AlCl₃ and 1 mg/kg of distilled water, orally for 2 weeks, sacrificed after 2wks), Group 3 (AlCl₃ .2wks)ii, received 100 mg/kg of AlCl₃ and 1 mg/kg of distilled water orally for 2 weeks (left untreated to check recovery), Group 4 (KS_{FLAV-MD} + AlCl₃) received 100 mg/kg of AlCl₃ + 200 mg/kg of flavonoids orally for 2 weeks, designated as medium dose, Group 5 (KS_{FLAV-HD} + AlCl₃) received 100 mg/kg of AlCl₃ + 300 mg/kg of flavonoids orally for 2 weeks, designated as High dose. After the administration, the animal sacrifice was done via cervical dislocation. Brain tissues were carefully harvested for homogenation assay while some were fixed in 10% neutral buffered formalin, processed and stained for histological studied. Results showed normal histoarchitecture of the cortical cells in group 1 (control) while the AlCl₃ treated group for 2 weeks and the group left untreated, revealed mild and marked distortion and cortical cells degeneration respectively with a significant (p < 0.05) depleted antioxidant enzymes (SOD, GSH, catalase) and elevated lipid peroxidation, MDA were observed compared to control. However, when treated with 200 mg/kg of KS flavonoid ((KS_{FLAV}- Medium dose) and 300 mg/kg of KS flavonoid (KS_{FLAV}- High dose) the results showed graded morphological neurorestorative process of the cortical cells, from minimal to normal orientation of the cortical cells and significantly restored antioxidant levels and reduced elevated lipid peroxidation by AlCl₃. Findings suggest that *Khaya senegalensis* flavonoids possess neurorestorative and anti-oxidant activities, which could be of potential benefit in the treatment and management of ischemic stroke.

Keywords: Ischemic Stroke, Parietotemporal Cortex, Aluminum Chloride, Flavonoid

Introduction

Medicinal plants have historically played a pivotal role in disease prevention and management, especially in developing countries where access to conventional pharmaceuticals may be limited. The World Health Organization (WHO) estimates that over 80% of the global population relies on traditional plant-based medicines for primary healthcare (World Health

Organization, 2013). This widespread use is attributed to the affordability, accessibility, and cultural acceptability of herbal remedies. Furthermore, the resurgence of interest in phytomedicine across modern science is driven by growing evidence that plant-based compounds possess a broad spectrum of pharmacological activities, including antioxidant, anti-inflammatory, antimicrobial, hepatoprotective, and neuroprotective effects (Cowan, 1999; Rates, 2001).

Khaya senegalensis, commonly known as African mahogany, is one such medicinal plant with significant pharmacological potential. Belonging to the family Meliaceae, this tree is widely distributed across tropical Africa, especially in West African countries like Nigeria, Senegal, Ghana, and Mali. Traditionally, the bark and leaves of *K. senegalensis* have been employed in the treatment of fever, malaria, gastrointestinal disorders, inflammation, and liver diseases (Okoye *et al.*, 2010). Phytochemical screening of the plant has revealed the presence of several bioactive constituents including flavonoids, tannins, alkaloids, saponins, steroids, triterpenoids, and limonoids—all of which contribute to its therapeutic properties (Akinmoladun *et al.*, 2007; Oboh *et al.*, 2016).

The neuroprotective potential of *Khaya senegalensis* is largely attributed to its high flavonoid content. Flavonoids have been shown to cross the blood-brain barrier, where they exert antioxidant and anti-apoptotic effects, enhance cerebral blood flow, reduce neuroinflammation, and modulate neuronal signaling cascades (Spencer, 2008). These mechanisms are vital in the context of ischemic stroke, where oxidative stress and inflammatory processes contribute significantly to neuronal damage. Given these properties, there is growing interest in exploring *K. senegalensis* as a candidate for neuroregenerative therapy in stroke models.

Among the classes of bioactive compounds found in plants, flavonoids are particularly significant. These polyphenolic secondary metabolites are present in almost all vascular plants and are known for their strong antioxidant properties. Flavonoids neutralize free radicals, chelate metal ions, modulate enzyme activity, and influence cellular signaling pathways (Middleton *et al.*, 2000). Their role in neuroprotection has received increasing scientific attention, as they have been shown to protect neurons from oxidative damage, inhibit neuroinflammation, and promote neurogenesis (Spencer, 2008). In light of this, medicinal plants rich in flavonoids are being evaluated as potential therapeutic agents for the management of stroke and neurodegenerative disorders such as Alzheimer's and Parkinson's diseases (Maher, 2019).

Stroke is a sudden and acute neurological event characterized by the rapid loss of brain function due to an interruption in the blood supply to the brain (Campbell *et al.*, 2019). The cessation of cerebral perfusion leads to deprivation of oxygen and glucose, which are critical for neuronal metabolism, resulting in irreversible brain tissue damage within minutes (Virani *et al.*, 2021).

Ischemic stroke is a major global health problem and ranks as the second leading cause of death and the third leading cause of disability worldwide (Feigin *et al.*, 2022). It results from an abrupt occlusion of cerebral blood vessels, leading to a reduction or complete cessation of

blood supply to parts of the brain. This disruption in cerebral circulation causes hypoxia, glucose deprivation, excitotoxicity, oxidative stress, and ultimately neuronal death (Dirnagl *et al.*, 1999). The parietotemporal cortex is one of the brain regions that is highly susceptible to ischemic damage due to its high metabolic activity and vascularization (Kumral *et al.*, 2002). Injury to this region often results in cognitive deficits, including impaired memory, visuospatial disorientation, and sensory integration dysfunctions (Corbetta & Shulman, 2002).

Although numerous pharmacological interventions exist for the management of stroke, such as thrombolytics and neuroprotective agents, their clinical utility is limited by narrow therapeutic windows, high cost, and adverse side effects (Powers *et al.*, 2018). Hence, there is a compelling need for affordable, safe, and effective alternatives—especially those derived from natural sources—that can mitigate stroke-related brain injury and enhance recovery outcomes.

In experimental models, neurotoxic agents like aluminum chloride (AlCl_3) are commonly used to simulate oxidative and ischemic brain injuries. Aluminum, a non-essential and neurotoxic metal, accumulates in the brain following chronic exposure and induces severe neuronal dysfunctions. Its neurotoxicity is mediated through several mechanisms including oxidative stress, mitochondrial dysfunction, inflammation, disruption of calcium homeostasis, and inhibition of essential enzymes such as acetylcholinesterase (Yokel & Florence, 2006; Exley, 2013). Studies have shown that AlCl_3 administration in laboratory animals leads to histopathological alterations in the cerebral cortex and hippocampus, mimicking features of stroke and neurodegeneration (Shati, 2011). As such, aluminum-induced models provide a useful platform for evaluating the efficacy of neuroprotective agents.

Given the known neurotoxic effects of aluminum Yokel & Florence, 2006; Exley, 2013 and the limited efficacy of conventional stroke therapies, it is both timely and necessary to explore alternative interventions. This research, therefore, seeks to evaluate the ameliorative effects of *Khaya senegalensis* flavonoids on aluminum chloride-induced ischemic stroke in the parietotemporal cortex of Wistar rats, using a combination of histological and biochemical techniques. The study aims to provide scientific insight into the potential role of *K. senegalensis* in neuroprotection, and by extension, contribute to the development of plant-based interventions for stroke management.

Materials and Method

Plant material

Fresh bark of *Khaya senegalensis* was gotten from a Forest at Ukele community of Cross River State. The tree bark was identified and authenticated in the Department of Botany, University of Lagos, Nigeria and stored in the herbarium; LUH 8004 - *Khaya senegalensis* (KS).

Animals

All protocols and treatment procedures of the experiment were approved by the Federal University Wukari Research Ethics Committee of the College of Health Sciences with protocol number FUW/CHS/HREC/JUNE/2025/015/VOL1. Total of 35 male Wistar rats weighing

150 - 220 g, were obtained from the animal house of College of Health Sciences, Federal University Wukari and were kept in well-ventilated plastic cages, kept and maintained under standard laboratory conditions in the animal house of the department of Anatomy of the faculty of Basic Medical Sciences of the Federal University Wukar. They were allowed free access to food and tap water ad libitum.

Materials and chemicals used

Cages, surgical gloves, cotton wool, face masks, spatula, oral cannula, feeding plates, distilled water, sensitive weighing balance, and web cam, syringe (1ml, 2ml, 5ml, and 10ml), feed, towel, mortar and pestle, normal saline, 10% formal-saline solution, phosphate-buffered saline (PBS), sample bottles, dissecting set.

Chemicals used

Aluminum chloride (AlCl₃) was obtained from the Histology laboratory of Department of Human Anatomy, Federal University Wukari, Wukari, Taraba state, Nigeria.

Extracts Preparation

Fresh bark of *Khaya senegalensis* was well cleansed and diced into smaller pieces using a sterile knife to aid the drying process and after which they were air dried at room temperature for a period of four weeks. The stem bark was then oven dried at 50°C for 3hrs and thereafter crushed into semi powder using a grinding machine. 244g of coarse powder of bark *Khaya senegalensis* was packed into a thimble and inserted to the Soxhlet extractor. The Soxhlet was inserted into the quick fit bottom flask containing solvent. The solution was left to concentrate using a rotary evaporator and the dried extract of *Khaya senegalensis* yielded 221g, was collected and preserved at 4°C for further use. And extraction of flavonoids was carried out according to Ushie *et al.*, 2022.

Dose Preparation of AlCl₃

The experimental rat's weights were measured and an average of 165.2g was taken. The dose of induction of the rats was calculated using the average weight of the experimental rats, the concentration of aluminum chloride dissolved in water.

Phytochemical Screening of *Khaya Senegalensis* bark

The phytochemical screening of the ethanolic extract was investigated using standard qualitative procedures (Trease & Evans, 1989; Sofowara, 1993) as reported by Kumar *et al.*, 2015; Lukpata *et al.*, 2020.

Experimental Design and Treatment

A total number of thirty - five healthy adults male wistar rats weighing 150 - 220 g, were used for this experiment and they were grouped into five groups ($n = 6$). The experiment was divided into 2 phases. Phase I: induction of experimental ischemic stroke, $AlCl_3$ was administered using a single dose of 0.5 mg/kg of $AlCl_3$ for 14 days and a control (CTL) group received 0.5 ML of normal saline for 28 days. Phase II: treatment groups, after induction of experimental stroke, the rats were divided into 3 subgroups; ($AlCl_3$ group (ischemic stroke rats), which was left untreated to check recovery, and two treatment groups, $KS_{FLAV-MD} + AlCl_3$ and $KS_{FLAV-HD} + AlCl_3$ groups) received 200 mg/kg and 300 mg/kg as medium and high doses respectively of KS_{FLAV} . Choice of dose selection was based on previous studies by Nwosu *et al.*, (2012), who reported that the LD₅₀ of the extract is greater than 3000mg/kg body weight.

Induction of Experimental Ischemic Stroke

$AlCl_3$ at a single dose of 0.5 mg/100g body weight was administered daily for 14 days. $AlCl_3$ was dissolved in 150 ML of water and dose was calculated by simple proportion based on animal weight and administered via oral route with the use of a metal oropharyngeal canula. Close daily food and water monitoring was done after $AlCl_3$ administration.

Dose treatment

Group 1: Control (received distilled water).

Group 2: $AlCl_3$ -2Wks(⁽ⁱ⁾) received 100 mg/kg body weight of $AlCl_3$ orally for 2 weeks, sacrificed after 2wks to evaluate ischemic stroke.

Group 3: $AlCl_3$ -2Wks(⁽ⁱⁱ⁾) (received 100 mg/kg body weight of $AlCl_3$ orally for 2 weeks, left untreated to check recovery).

Group 4: $KS_{FLAV-MD} + AlCl_3$ (received 100 mg/kg body weight of $AlCl_3$ + 200 mg/kg of flavonoids orally for 2 weeks, designated as medium dose).

Group 5: $KS_{FLAV-HD} + AlCl_3$ (received 100 mg/kg body weight of $AlCl_3$ + 300 mg/kg of flavonoids orally for 2 weeks, designated as High dose).

Acute Toxicity Test

The acute toxicity effect of the extract was determined using the fixed dose protocol of the Organization of Economic Co-operation and Development (OECD) guidelines for testing of chemicals, TG420 (OECD & Staff, 2001) for oral administration.

Doses of *khaya senegalensis* flavonoid extract and Aluminum Chloride ($AlCl_3$) was determined based on prior toxicity and efficacy studies, which ensured the safety and pharmacological relevance of treatment regimens. (Agyare *et al.* 2013). In the case of aluminum chloride, acute toxicity studies in rodents have shown that the oral LD₅₀ is approximately 3,311–3,470 mg/kg body weight in rats, indicating that it has moderate acute toxicity when ingested (Fisher Scientific, 2020; Ricca Chemical, 2025; Sigma-Aldrich, 2024; Spectrum Chemical, 2020).

Conversely, acute toxicity studies on the aqueous stem bark extract of *Khaya senegalensis* reveal an LD₅₀ greater than 5,000 mg/kg body weight in rats, with no significant adverse clinical or histopathological changes, suggesting that the bark extract is relatively safe at therapeutic doses (Akintola *et al.*, 2013)

Dose Standardization

For the treatment group; each wistar rat was treated with *Khaya senegalensis* flavonoid, group 4; medium dose of *khaya senegalensis*, group 5; high dose of *khaya senegalensis* flavonoid, received 200mg/kg and 300mg/kg per body weight as medium and high doses of *Khaya senegalensis* once a day for a period of fourteen (14) days.

Animal Sacrifice

The animal sacrifice was done via cervical dislocation. Subsequently, the skull was opened to harvest the whole brain which was carefully divided into two through the longitudinal fissure one part was fixed in the perfusion fixative (10% formal saline) and the other part was homogenized with mechanical homogenizer in a Phosphate buffer saline solution and collected in a plain tube. Subsequently the homogenized brain tissue was centrifuged at 3500rpm to separate debris or insoluble materials, the resultant supernatant was taken for biochemical analysis. Afterwards, for routine histological processing was carried out in the Neuro-Lab, Akure Ondo state Nigeria, while biochemical analysis was done at Lively Stone Lab, Port-Harcourt Rivers state Nigeria. After staining with H and E microscopic slides where viewed under the microscope using X400 and photomicrographs where taken using MD900 Amscope® digital camera.

Results

Biochemical assay: in our study the control group demonstrated normal antioxidant balance with significantly higher activities of superoxide dismutase (SOD), glutathione (GSH), and catalase, alongside a low malondialdehyde (MDA) level. In contrast, the result of the Oxidative Stress marker revealed a significantly decrease ($p<0.05$) in antioxidant activities; superoxide dismutase (SOD), glutathione (GSH), and catalase (CAT) and significantly ($p<0.05$) increase in MDA level in the group that received 100 mg/kg body weight of AlCl₃ orally for 2 weeks, sacrificed after 2wks (AlCl₃-2Wks(i)). In AlCl₃-2Wks(ii) group which received 100 mg/kg body weight of AlCl₃ orally for 2 weeks, and left untreated for 2 weeks to check recovery revealed that after 2 weeks, there was partial improvement in SOD and MDA values, although catalase remained severely diminished, indicating sustained oxidative burden and potential compensatory mechanisms. The administration of *Khaya senegalensis* flavonoids in AlCl₃-treated rats produced notable ameliorative effects. At medium dose, there was a mild increase in SOD and catalase compared to the AlCl₃ groups, and MDA levels were reduced, suggesting a degree of neuroregeneration. On treatment with KS_{FLAV-HD} shows a significant improvement in antioxidant enzyme activities and a further reduction in MDA, reflecting more restorative effect.

Table 1: Effect administration aluminium chloride and *Khaya senegalensis* flavonoid on Oxidative Stress marker

Groups	SOD(U/ml)	GSH (mmol)	MDA (umol)	Catalase (u/ml)
Control	4.11±0.12	1.79±0.25 ^a	0.86±0.12 ^a	198.99±47.09 ^a
AlCl ₃ -2Wks(i)	0.27±0.03*	0.18±0.04*	5.00±0.27*	56.92±7.43*
AlCl ₃ -2Wks(ii)	1.28±0.01* ^a	0.44±0.04*	1.27±0.18 ^a	12.21±0.01*
KS _{FLAV-MD} + AlCl ₃	1.03±0.02* ^a	0.35±0.01*	1.22±0.01 ^a	8.76±0.04*
KS _{FLAV-HD} + AlCl ₃	0.83±0.02* ^a	0.50±0.03*	1.27±0.12 ^a	4.84±0.02*

n=3; mean ± SEM, one-way ANOVA, * = p<0.05; significance difference when compared to control; a=p<0.05 when compared to AlCl₃ (2Wks).

Histological studies: In fig Figure 1, A photomicrograph of a section in the parieto-temporal cortex), Control (CTL) received distilled water, showing a normal architecture of cerebral cortex, featuring the pyramidal cell, Neuroglial cell, With dense nuclei Granule cell, With pale open face nucleus. AlCl₃ treated group which received 100 mg/kg body weight of AlCl₃ orally for 2 weeks, sacrificed after 2wks (AlCl₃ -2Wks(i)), revealed cortical cells degeneration characterized by astrocytosis, Lymphocytes Infiltrates and Vacuolated Neutrophils, pyramidal cell with irregular shape and surrounded by pericellular halos, Perivascular Edema, Perivascular Cuffing and shrunken Granule cells deeply stained. AlCl₃ -2Wks(ii) group which received 100 mg/kg body weight of AlCl₃ orally for 2 weeks, and left untreated for 2 weeks to check recovery, showed marked cortical cells degeneration characterized by astrocytosis, degenerate pyramidal neuron with irregular shape and surrounded by pericellular halos, Perivascular Edema, Perivascular Cuffing and shrunken Granule cells deeply stained. KS_{FLAV-MD} + AlCl₃ treated rats showed less or normal neurovascular unit, Pyramidal cell, Neuroglial cells and Neutrophils vacuolated, Granule cells. Pyramidal cell, Neuroglial cells and, Granule cells (GC) visibly showed open face nuclei and basophil cytoplasm, prominent nucleoli. KS_{FLAV-HD} + AlCl₃ treated rats. Showing more prominent orientation of blood vessels, Pyramidal cell, Neuroglial cells and Neutrophils vacuolated, Granule cells. Pyramidal cell, Neuroglial cells darkly stained and Granule cells visibly showing open face nuclei and basophil cytoplasm, prominent nucleoli.

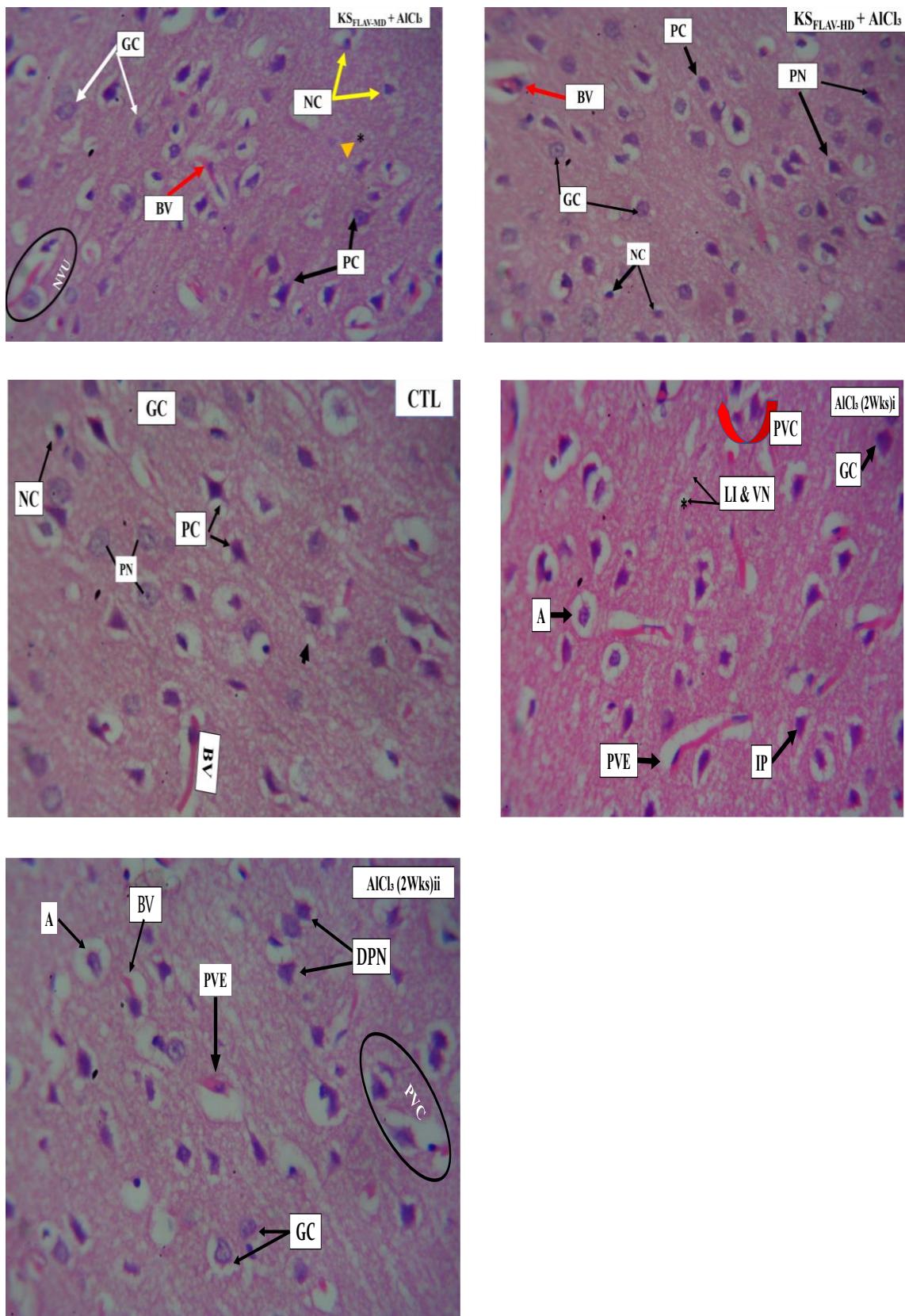


Fig 1: A photomicrograph of a section in the parieto-temporal cortex of male Wister rats. Control (CTL) showing normal architecture of cerebral cortex, pyramidal cell (P), Neuroglial cell (N), With dense nuclei granule cell (G), AlCl₃ -2Wks(i) showing mild cortical cells degeneration characterized by astrocytosis (A), Lymphocytes Infiltrates (LI) and Vacuolated Neutrophils (VN), pyramidal cell with irregular shape (IP) and surrounded by pericellular halos, Perivascular Edema (PVE), Perivascular Cuffing (PVC) and shrunken Granule cells (GC). AlCl₃ -2Wks(ii) showing marked cortical cells degeneration characterized by astrocytosis(A), degenerate pyramidal neuron with irregular shape (DPN) and surrounded by pericellular halos, Perivascular Edema (PVE), Perivascular Cuffing (PVC) and shrunken Granule cells (GC). KSFLAV-MD + AlCl₃ treated rats, Showing less or normal neurovascular unit (NVN), Pyramidal cell (PC), Neuroglial cells (NC) and Neutrophils (N) vacuolated, Granule cells (GC). Pyramidal cell (PC), Neuroglial cells (NC) and, Granule cells (GC) visibly showing open face nuclei and basophil cytoplasm, prominent nucleoli. KSFLAV-HD + AlCl₃ treated rats. Showing more prominent orientation of blood vessels (BV), Pyramidal cell (PC), Neuroglial cells (NC) and Neutrophils (N) vacuolated, Granule cells (GC). Pyramidal cell (PC), Neuroglial cells (NC) darkly stained and Granule cells (GC) visibly showing open face nuclei and basophil cytoplasm, prominent nucleoli. (H & E X400)

Discussion

In the past years, Cerebrovascular accident (CVA) is considered to be the third most common cause of mortality in the developed world. Current treatments such as tissue plasminogen activator (tPA/Alteplase) is the gold standard for acute ischemic stroke if administered within 4.5 hours, but its effectiveness declines rapidly with delayed administration, with high risk of intracerebral hemorrhage (about 6%), not to mention systemic bleeding and other complications (Hacke *et al.*, 2008).

In our present study, the biochemical analysis of oxidative stress biomarkers is presented in Table 1. The control group demonstrated normal antioxidant balance with significantly higher activities of superoxide dismutase (SOD), glutathione (GSH), and catalase, alongside a low malondialdehyde (MDA) level, reflecting a healthy redox state. This finding aligns with earlier reports that physiological antioxidant systems maintain cellular homeostasis under normal conditions (Halliwell & Gutteridge, 2015). In contrast, aluminum chloride (AlCl₃) administration for two weeks caused a marked depletion of antioxidant defenses, as indicated by significantly decreased SOD, GSH, and catalase levels, while MDA was drastically elevated compared to the control ($p < 0.05$). This observation is consistent with previous studies that demonstrated AlCl₃-induced oxidative damage in neuronal tissues through lipid peroxidation and suppression of endogenous antioxidants (Ebhodaghe *et al.*, 2019; Suryavanshi *et al.*, 2022). In AlCl₃-2Wks(ii) group which received 100 mg/kg body weight of AlCl₃ orally for 2 weeks, and left untreated for 2 weeks to check recovery, there was partial improvement in SOD and MDA values, although catalase remained severely diminished, indicating sustained oxidative burden and potential compensatory mechanisms. The administration of Khaya senegalensis flavonoids in AlCl₃-

treated rats produced notable ameliorative effects. At medium dose, there was a modest increase in SOD and catalase compared to the $AlCl_3$ groups, and MDA levels were reduced, suggesting a degree of neuroprotection. At high dose, flavonoid treatment resulted in greater improvements in antioxidant enzyme activities and a further reduction in MDA, reflecting a stronger restorative effect. These findings corroborate earlier reports that flavonoids exert antioxidant and neuroprotective effects by scavenging free radicals, enhancing endogenous enzymatic defenses, and reducing lipid peroxidation (Spencer, 2008; Oyelami *et al.*, 2018). Specifically, extracts of *Khaya senegalensis* have been documented to contain polyphenolic compounds with potent antioxidant capacity, which contribute to protection against chemically induced oxidative insults (Abdullahi *et al.*, 2019; Ibrahim *et al.*, 2021).

The present study histological findings of parieto-temporal cortex in group 1 (control) showed normal histoarchitecture of the cortex with normal pyramidal cell, neurovascular unit, neuron, blood vessel, neuroglial cell with dense nuclei, granule cell, pale open face nucleus and blood vessels. This is in line with the work done by Ebhodaghe & Enogieru, 2024. Also, in the $AlCl_3$ treated groups for 2 weeks, showed mild and marked cortical cells degeneration characterized by astrocytosis, Lymphocytes Infiltrates and Vacuolated Neutrophils, pyramidal cell with irregular shape and surrounded by pericellular halos, Perivascular Edema, Perivascular Cuffing and shrunken Granule cells deeply stained. This suggest that there was neurodegeneration. The result of our present is consistent with the study Okhah and Enogieru (2023) who observed that, atrophy and vacuolation of astrocytes and pyramidal cells were seen.

On treatment, it was observed in this study that, the parieto-temporal cortex of $AlCl_3$ and KS_{FLAV} treated rats ($KS_{FLAV-MD} + AlCl_3$) unveils less or normal neurovascular unit, Pyramidal cell, Neuroglial cells and Neutrophils vacuolated, Granule cells. Pyramidal cell, Neuroglial cells and, Granule cells (GC) visibly showed open face nuclei and basophil cytoplasm, prominent nucleoli. This suggests that low dose (200 mg/kg) of *khaya Senegalensis* and have low ameliorative effect on aluminum chloride ($AlCl_3$)-induced stroke. This study correlates with Onu *et al.*, (2013) who also observed that for *khaya Senegalensis* is dose dependent.

In addition, on treatment with 300 mg/kg of *khaya senegalensis* on aluminum chloride ($AlCl_3$)-induced stroke in the parieto-temporal cortex of adult wistar rats ($KS_{FLAV-HD} + AlCl_3$). It reveals the ameliorative effects of *khaya Senegalensis* aluminum chloride ($AlCl_3$)-induced stroke which fully unveils the in the parieto-temporal cortex of $KS_{FLAV-HD} + AlCl_3$ treated rats. Showing more prominent orientation of blood vessels, Pyramidal cell, Neuroglial cells and Neutrophils vacuolated, Granule cells. Pyramidal cell, Neuroglial cells darkly stained and Granule cells visibly showing open face nuclei and basophil cytoplasm, prominent nucleoli.

This result implies that *khaya Senegalensis* 300 mg/kg have the efficacy to curtail and eradicate the toxic effect of induced stroke by aluminum chloride. This is in line with Onu *et al.*, (2013) who suggest that the aqueous stem bark extract of *K. senegalensis* may affect the cellular integrity of vital organs of the body.

Conclusion

In this our study, and from the results obtained above, *K_{sf}FLAV* have the efficacy and potency to ameliorate aluminum chloride (AlCl₃)-induced ischemic stroke on the parieto-temporal cortex. The result showed that at 100 mg/kg of aluminum chloride, stroke can be induced in fourteen days.

In conclusion, *Khaya senegalensis* flavonoids at dose 200 mg/kg have shown neuroregenerative and neurorepairs role on neurodegenerative changes in the parieto-temporal cortex, by regenerating pyramidal cell/granule cells and neurovascular unit.

Recommendation to Further Study

Additional studies involving different animal models and prolonged treatment periods should be conducted to establish optimal dosage, safety profiles, and possible toxicological limits before progressing to human clinical trials. Also, the compatibility of this herb and any other drug that induce stroke is recommended.

REFERENCES

Abdullahi M. S., Abubakar K., & Ahmed A. (2019). Phytochemical screening and antioxidant activities of *Khaya senegalensis* stem-bark extract. *Journal of Medicinal Plants Research*, 13(7), 164–170.

Abubakar A. R., & Haque M. (2015). Ethnopharmacology of *Khaya senegalensis*. *Journal of Ethnopharmacology*, 168, 361–368.

Adams H. P., del Zoppo G., Alberts M. J., Bhatt D. L., Brass L., Furlan A., ... & American Heart Association. (2007). Guidelines for the early management of adults with ischemic stroke. *Stroke*, 38(5), 1655–1711.

Adedapo A. A., Jimoh F. O., Koduru S., Masika P. J., & Afolayan A. J. (2014). Essential oil composition of *Khaya senegalensis* leaves. *Natural Product Communications*, 9(6), 869–872.

Adesokan A. A., Yakubu M. T., Owoyele B. V., Akanji M. A., & Soladoye A. O. (2008). Anti-inflammatory effect of *Khaya senegalensis* stem bark extract in rats. *Journal of Ethnopharmacology*, 120(2), 283–286.

Aebi, H. (1984). Catalase in vitro. *Methods in Enzymology*, 105, 121–126.

Akinmoladun A. C., Ibukun, E. O., Afor E., Obuotor E. M., & Farombi E. O. (2007). Phytochemical constituents and antioxidant activity of extract from the leaves of *Ocimum gratissimum*. *Scientific Research and Essay*, 2(5), 163–166.

Akintola, A. O., Kehinde, A. O., & Olukayode, A. A. (2013). Effect of aqueous stem bark extract of *Khaya senegalensis* on biochemical, haematological, and histopathological parameters in rats. *Nigerian Journal of Physiological Sciences*, 28(1), 65–71.

An J., Li, H., Tang Y., Zheng N., Ye, C., Wu Y., ... & Yang G. (2017). Intracerebral hemorrhage: Pathophysiology and management. *CNS Neuroscience & Therapeutics*, 23(6), 476–484.

Atawodi S. E., Bulus T., Ibrahim S., Ameh D. A., Nok A. J., Mamman M., & Galadima M. (2010). *In vitro* antitrypanosomal activity of extracts of *Khaya senegalensis*. *African Journal of Biotechnology*, 9(42), 6912–6916.

Balkaya M., Kröber J. M., Rex A., & Endres M. (2013). Assessing post-stroke behavior in mouse models of focal ischemia. *Journal of Cerebral Blood Flow & Metabolism*, 33(3), 330–338.

Bamford J., Sandercock P., Dennis M., Burn J., & Warlow, C. (1991). Classification and natural history of clinically identifiable subtypes of cerebral infarction. *The Lancet*, 337(8756), 1521–1526.

Bondy S. C. (2010). The neurotoxicity of environmental aluminum is still an issue. *NeuroToxicology*, 31(5), 575–581.

Burkill H. M. (1985). *The useful plants of West Tropical Africa* (Vol. 1). Royal Botanic Gardens, Kew.

Campbell B. C. V., De Silva D. A., Macleod M. R., Coutts, S. B., Schwamm L. H., Davis S. M., & Donnan, G. A. (2019). Ischaemic stroke. *Nature Reviews Disease Primers*, 5(1), 70.

Caplan L. R. (2000). Top of the basilar syndrome. *Neurology* 55(3), 462–463.

Cao Z., Yang X., Zhang H., Wang H., Huang W., Li, T., ... & Wang T. (2016). Aluminum chloride induces neuroinflammation via the NLRP3 inflammasome activation. *Journal of Alzheimer's Disease*, 52(1), 1–15.

Caspers S., Geyer S., Schleicher A., Mohlberg H., Amunts K., & Zilles K. (2006). The human inferior parietal cortex: Cytoarchitectonic mapping and interindividual variability. *NeuroImage*, 33(2), 430–448.

Chen L., Yokel R. A., Hennig B., & Toborek M. (2008). Manufactured aluminum oxide nanoparticles decrease expression of tight junction proteins in brain vasculature. *Journal of Neuroimmune Pharmacology*, 3(4), 286–295.

Connolly J. D., Labbé C., Rycroft D. S., & Karchesy J. J. (1999). Limonoids of *Khaya senegalensis*. *Phytochemistry*, 52(6), 1105–1110.

Corbetta M., Kincade M. J., Lewis C., Snyder A. Z., & Sapir A. (2005). Neural basis and recovery of spatial attention deficits in spatial neglect. *Nature Neuroscience*, 8(11), 1603–1610.

Corbetta M., & Shulman G. L. (2002). Control of goal-directed and stimulus-driven attention in the brain. *Nature Reviews Neuroscience*, 3(3), 201–215.

Delshadi R., Bahrami A., Assadpour E., & Jafari S. M. (2019). *Potential inhibitory effect of natural compounds against platelet-activating factors in ischemic stroke*. *Critical Reviews in Food Science and Nutrition*, 59(15), 2441–2468.

Delshadi R., Bahrami A., Assadpour E., & Jafari S. M. (2019). *Potential inhibitory effect of natural compounds against platelet-activating factors*

in ischemic stroke. Critical Reviews in Food Science and Nutrition, 59(15), 2441–2468.

Dirnagl U., Iadecola C., & Moskowitz M. A. (1999). Pathobiology of ischaemic stroke: An integrated view. *Trends in Neurosciences, 22*(9), 391–397.

Easton J. D., Saver J. L., Albers G. W., Alberts M. J., Chaturvedi S., Feldmann E., ... & American Heart Association. (2009). Definition and evaluation of transient ischemic attack. *Stroke, 40*(6), 2276–2293.

Ebhodaghe C. I., Akinmoladun F. O., Komolafe T. R., Olaleye T. M., & Farombi O. E. (2019). Aluminium chloride-induced cerebral toxicity in Wistar rats: Anticholinesterase and antioxidant effects of ascorbic acid. *Journal of Chemical Neuroanatomy, 98*, 27–34.

Ebhodaghe C. I., & Enogieru A. B. (2024). Aluminium chloride-induced cerebral toxicity in Wistar rats: Anticholinesterase and antioxidant effects of ascorbic acid. *Journal of Phytomedicine and Therapeutics, 23*(2), 1548–1556.

Ekor M. (2014). The growing use of herbal medicines. *Frontiers in Pharmacology, 4*, 177.

Ellman, G. L. (1959). Tissue sulfhydryl groups. *Archives of Biochemistry and Biophysics, 82*(1), 70–77.

Exley C. (2013). Human exposure to aluminium. *Environmental Science: Processes & Impacts, 15*(10), 1807–1816.

Feigin V. L., Stark B. A., Johnson C. O., Roth G. A., Bisignano C., Abady G. G., ... & Murray C. J. L. (2022). Global, regional, and national burden of stroke and its risk factors, 1990–2019: A systematic analysis for the Global Burden of Disease Study 2019. *The Lancet Neurology, 20*(10), 795–820.

Fisher Scientific. (2020). *Safety Data Sheet: Aluminum chloride hexahydrate*. Fisher Scientific.

Food and Agriculture Organization (FAO). (1986). *Khaya senegalensis* (Desr.) A. Juss. *Forestry Paper*.

Gill L. S. (1992). *Ethnomedical uses of plants in Nigeria*. Uniben Press.

Global Burden of Disease 2021 Stroke Collaborators. (2023). Global stroke statistics 2021. *The Lancet Neurology, 22*(1), 18–32.

Hacke W., Kaste M., Bluhmki E., Brozman M., Dávalos A., Guidetti D., Larrue V., Lees K. R., Medeghri Z., Machnig T., Schneider, D., von Kummer R., Wahlgren N., & Toni D. (2008). Thrombolysis with alteplase 3 to 4.5 hours after acute ischemic stroke. *The New England Journal of Medicine, 359*(13), 1317–1329.

Halliwell, B., & Gutteridge, J. M. C. (2015). Free radicals in biology and medicine (5th ed.). Oxford University Press.

Hemphill J. C., Greenberg S. M., Anderson C. S., Becker K., Bendok B. R., Cushman M., ... & American Heart Association Stroke Council. (2015). Guidelines for the management of spontaneous intracerebral hemorrhage. *Stroke, 46*(7), 2032–2060.

Hillis A. E. (2007). Aphasia: Progress in the last quarter of a century. *Neurology, 69*(2), 200–213.

Hurd M. D., Norrvig, B., & Ali, M. (2021). *Combined antiplatelet and thrombolytic therapy in acute ischemic stroke: Current perspectives.* *Journal of Thrombosis and Haemostasis*, 19(5), 1175–1185.

Ibrahim, M., Okonji, R. E., & Suleiman, A. (2021). Antioxidant and anti-inflammatory activities of *Khaya senegalensis* stem bark extract in oxidative stress models. *African Journal of Traditional, Complementary and Alternative Medicines*, 18(2), 34–42.

Ibrahim H., Musa, A., Aliyu A. B., & Mayaki H. S. (2013). *In vitro* antiplasmodial activity of *Khaya senegalensis* extracts. *Journal of Acute Disease*, 2(4), 285–288.

Isa A. I., Awouafack M. D., Dzoyem J. P., Aliyu M., Magaji R. A., & Ayo, J. O. (2015). Morphological diversity in *Khaya senegalensis* populations. *Journal of Forestry Research*, 26(3), 617–624.

Iwu M. M. (1993). *Handbook of African medicinal plants*. CRC Press.

Kleindorfer D. O., Towfighi A., Chaturvedi S., Cockroft K. M., Gutierrez J., Lombardi-Hill D., ... & Williams L. S. (2021). 2021 Guideline for the prevention of stroke in patients with stroke and transient ischemic attack. *Stroke*, 52(7), e364–e467.

Koné W. M., Atindehou K. K., Terreaux C., Hostettmann K., Traoré D., & Dosso M. (2012). Ethnobotanical study of plants used to treat diabetes. *Journal of Ethnopharmacology*, 143(1), 367–371.

Kumral E., Bayulkem G., Evyapan D., & Yunten N. (2002). Spectrum of anterior cerebral artery territory infarction: Clinical and MRI findings. *European Journal of Neurology*, 9(6), 615–624.

Kumar V., Bal A., & Gill, K. D. (2009). Aluminum-induced oxidative stress in rat brain. *Neurotoxicology*, 30(5), 752–762.

Langhorne P., Bernhardt J., & Kwakkel G. (2011). Stroke rehabilitation. *The Lancet*, 377(9778), 1693–1702.

Lukpata PU, Gbotolorun SC, Oremosu AA, & Tokede OE. (2020). Ethanolic Stem Bark Extract of *Khaya Senegalensis* Ameliorates Cerebral Ischemia in Wistar Rats. *Journal of Anatomical Sciences*, 11(2), (41-49).

Maher P. (2019). The potential of flavonoids for the treatment of neurodegenerative diseases. *International Journal of Molecular Sciences*, 20(12), 3056.

Maya S., Prakash T., Madhu K. D., & Goli D. (2016). Multifaceted effects of aluminium in neurodegenerative diseases: A review. *Biomedicine & Pharmacotherapy*, 83, 746–754.

McCrory M. R., van der Worp, H. B., & Bath, P. M. (2017). *Adjunctive antiplatelet therapy for reperfusion in ischemic stroke: A meta-analysis.* *Stroke*, 48(7), 1877–1883.

Meschia J. F., Bushnell C., Boden-Albala B., Braun L. T., Bravata D. M., Chaturvedi S., ... & Wilson J. A. (2014). Guidelines for the primary prevention of stroke. *Stroke*, 45(12), 3754–3832.

Middleton E., Kandaswami C., & Theoharides T. C. (2000). The effects of plant flavonoids on mammalian cells: Implications for inflammation, heart disease, and cancer. *Pharmacological Reviews*, 52(4), 673–751.

Misra H. P., & Fridovich I. (1972). The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. *Journal of Biological Chemistry*, 247(10), 3170–3175.

Newton A. C., Baker P., Ramnarine S., Mesén J. F., & Leakey R. R. B. (1993). The mahogany shoot borer: Prospects for control. *Forest Ecology and Management*, 57(1-4), 301–328.

Noumi E., & Yomi A. (2001). Medicinal plants used for intestinal diseases in Mbalmayo Region, Central Province, Cameroon. *Fitoterapia*, 72(3), 246–254.

Nwidu L. L., Elmorsy E., Thornton J., & Wijamunige B. (2015). Neuroprotective effects of *Khaya senegalensis* in scopolamine model. *Journal of Ethnopharmacology*, 172, 325–334.

Oboh G., Ademosun A. O., & Ogunsuyi O. B. (2016). Antioxidant and anticholinesterase activities of extract from *Hibiscus sabdariffa* leaves: Impact of ripening. *Journal of Food Biochemistry*, 40(3), 275–282.

Ohkawa H., Ohishi N., & Yagi K. (1979). Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Analytical Biochemistry*, 95(2), 351–358.

Onu A., Saidu Y., Ladan M. J., Bilbis L. S., Aliero A. A., & Sahabi S. M. (2013). Effect of aqueous stem bark extract of *Khaya senegalensis* on some biochemical, haematological, and histopathological parameters of rats. *Journal of toxicology*, 2013.

O'Donnell M. J., Chin S. L., Rangarajan S., Xavier D., Liu L., Zhang H., ... & INTERSTROKE Investigators. (2016). Global and regional effects of potentially modifiable risk factors associated with acute stroke in 32 countries (INTERSTROKE). *The Lancet*, 388(10046), 761–775.

Okhah A. A., & Enogieru A. B. (2023). Antioxidant and protective activities of aqueous *Theobroma cacao* seed extract against aluminium-induced hippocampal toxicity in Wistar rats. *Nigerian Journal of Biochemistry and Molecular Biology*, 38(4).

Okoh O. O., Nwodo U. U., Ibezim A., Noundou X. S., & Okeke I. N. (2015). Neuroprotection by *Terminalia catappa* in ischemia-reperfusion. *Pharmaceutical Biology*, 53(12), 1779–1789.

Okorie O. K., Akah P. A., Okoli C. O., & Ezeugwu N. (2016). Hepatoprotective effects of *Khaya senegalensis* bark extract. *Pharmaceutical Biology*, 54(10), 2107–2113.

Okoye T. C., Akah P. A., & Okoli C. O. (2010). *Khaya senegalensis* stem bark extract protects against acute experimental inflammation and chemical-induced arthritis in rats. *Pharmaceutical Biology*, 48(8), 885–891.

Orwa C., Mutua A., Kindt R., Jamnadass R., & Simons A. (2009). *Khaya senegalensis*. In *Agroforestry Database: A tree reference and selection guide version 4.0*. World Agroforestry Centre.

Ovbiaghe B., & Goldstein L. B. (2014). Primary prevention of stroke. *Circulation Research*, 114(7), 1124–1129.

Owolabi M. O., Akarolo-Anthony S., Akinyemi R., Arnett D., Gebregziabher M., Jenkins C., ... & Stroke Investigators. (2015). The burden of stroke in Africa. *International Journal of Stroke*, 10(SA100), 1–3.

Oyelami T. A., Adeleye A. O., & Oladimeji S. A. (2018). Neuroprotective role of flavonoids in oxidative stress-related neurodegeneration. *Nigerian Journal of Physiological Sciences*, 33(2), 105–112.

Pan R., Qiu S., Lu D. X., & Dong J. (2012). Curcumin attenuates neuroinflammation in focal cerebral ischemia. *Journal of Cerebral Blood Flow & Metabolism*, 32(7), 1332–1345.

Peters S. A. E., Muntner P., & Woodward M. (2019). Alcohol consumption and risk of stroke. *Stroke*, 50(2), 531–535.

Powers W. J., Rabinstein A. A., Ackerson T., Adeoye O. M., Bambakidis N. C., Becker K., ... & American Heart Association Stroke Council. (2018). Guidelines for the early management of patients with acute ischemic stroke. *Stroke*, 50(12), e344–e418.

Prakash A., Kumar A., & Kumar V. (2013). Aluminum-induced oxidative stress and mitochondrial dysfunction are mitigated by *Withania somnifera* via NF-κB regulation. *Journal of Alzheimer's Disease*, 37(1), 143–153.

Qureshi A. I., Tuhrim S., Broderick J. P., Batjer H. H., Hondo H., & Hanley D. F. (2001). Spontaneous intracerebral hemorrhage. *New England Journal of Medicine*, 344(19), 1450–1460.

Rates S. M. K. (2001). Plants as source of drugs. *Toxicon*, 39(5), 603–613.

Ricca Chemical Company. (2025). *Safety Data Sheet: Aluminum chloride hexahydrate*. Ricca Chemical

Saha S., & Hossain M. S. (2022). Aluminum chloride: Properties, applications, and safety. *ChemistrySelect*, 7(12), e202104046.

Sands M. J. S. (2001). *The desertification of the West African Sahel*. University of Oxford Press.

Seghier M. L. (2013). The angular gyrus: Multiple functions and multiple subdivisions. *The Neuroscientist*, 19(1), 43–61.

Shati A. A. (2011). Effects of *Morus alba* leaves extract on the brain of male rats exposed to aluminum chloride. *Journal of American Science*, 7(2), 377–385.

Sidiyasa K. (1998). Taxonomy, phylogeny, and wood anatomy of *Khaya* (Meliaceae). *Blumea*, 43(1), 1–144.

Sigma-Aldrich. (2024). *Safety Data Sheet: Aluminum chloride, anhydrous*. Sigma-Aldrich.

Sofowora A. (2008). *Medical plants and traditional medicine in Africa* (3rd ed.). Spectrum Books.

Spectrum Chemical. (2020). *Material Safety Data Sheet: Aluminum chloride, hexahydrate*. Spectrum Chemical.

Spencer J. P. E. (2008). Flavonoids: Modulators of brain function? *British Journal of Nutrition*, 99(E-S1), ES60–ES77.

Suryavanshi J., Prakash C., & Sharma D. (2022). Asiatic acid attenuates aluminium chloride-induced behavioral changes, neuronal loss and astrocyte activation in rats. *Metabolic Brain Disease*, 37(6), 1773–1785.

Teklehaimanot Z., Tomlinson H., & Reeves C. (2004). *Khaya senegalensis* in agroforestry systems. *Forest Ecology and Management*, 189(1-3), 437–446.

Ugbabe G. E., Ezeunala M. N., Edmond I. N., Amedu N. O., & Salaam A. (2010). Preliminary phytochemical and antimicrobial screening of *Khaya senegalensis* leaves. *Journal of Medicinal Plants Research*, 4(10), 981–984.

Ushie O. A., Aikhoje E. F., Aasegh T. J., Ibrahim A. I., Bako B. & Ettah A. O. (2022). Estimation of total Alkaloids, Saponins Flavonoid, Tannins and Phenols in Thaumatococcus Danielli Leaves. *World journal of pharmaceutical and medical research*, 8(4), 242-246.

van Gijn, J., & Rinkel, G. J. E. (2001). Subarachnoid haemorrhage: Diagnosis, causes and management. *Brain*, *124*(2), 249–278.

Virani S. S., Alonso A., Aparicio H. J., Benjamin E. J., Bittencourt M. S., Callaway C. W., ... & Stroke Statistics Subcommittee. (2021). Heart disease and stroke statistics—2021 update. *Circulation*, 143(8), e254–e743.

Wei Y., Zhang T., & Ito Y. (2020). *Natural antiplatelet agents from botanical sources for ischemic stroke therapy*. *Journal of Ethnopharmacology*, 252, 112591.

World Health Organization. (2013). WHO traditional medicine strategy: 2014-2023. World Health Organization.

Yakubu M. T., & Bukoye B. B. (2009). Reproductive toxicity of aqueous extract of *Khaya senegalensis* stem bark in male rats. *Journal of Ethnopharmacology*, 126(3), 507–513.

Yakubu M. T., Quadri A. L., & Owoyele B. V. (2013). Neuroprotective effects of *Khaya senegalensis* in aluminum-induced neurodegeneration. *Pharmaceutical Biology*, 51(1), 52–60.

Yokel R. A., & Florence R. L. (2006). Aluminum bioavailability from tea infusion. *Food and Chemical Toxicology*, 44(11), 1902–1912.

Zinkstok S. M., Roos Y. B., & ARTIS Investigators. (2010). *Antiplatelet therapy in combination with rt-PA thrombolysis in ischemic stroke (ARTIS): Rationale and design of a randomized controlled trial*. *Cerebrovascular Diseases*, 29(1), 79–81.

Zhou L., Li, F., Xu H., Luo C., & Xia Q. (2006). *Ginkgo biloba* extract reduces infarct volume in rat middle cerebral artery occlusion model. *Journal of Ethnopharmacology*, 107(2), 242–250.